

Combined effect of AlCl₃ and GA on Biochemical Characterization of Wheat Primary Leaves during Dark Incubated Senescence

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Abstract

Senescence is the age-dependent end of the life span. In plants, it can be visualized by yellowing of leaves that accompanies the mobilization of leaf nutrients to the reproductive structures. The yellowing of senescing leaves is correlated with a series of biochemical changes such as loss of chlorophyll contents, degradation of proteins, RNA and a decline in photosynthetic activity. Senescence limits crop yields in annual crops like maize, rice and wheat. Delayed leaf senescence is a desirable agronomic trait to improve crop yield. In this study the combination of AlCl₃ and Gibberellic acid reduced the loss of pigments, proteins, spectral properties, electron transport activities in wheat primary leaves under incubated dark conditions. The restoration of whole chain electron transport activity by the combination of AlCl₃ and GA was closely associated with the restoration of PS II activity when compared with PS I. The combination of Gibberellic acid and AlCl₃ treated leaf thylakoid membranes showed an increase in absorption at 680 nm moderate increases at 480 nm and 440 nm at 72 h during dark incubation. The AlCl₃ and Gibberellic acid protected the degradation of water oxidation complex polypeptides (33, 23, 17 KDa) of PS II and slightly protected the 68 kDa of PS I polypeptides.

Keywords: Senescence retardation; PSII and PS I activity; water oxidation complex polypeptides

Introduction

Senescence is defined as a time-dependent, gradual decay of multiple biological functions (López -Otín et al., 2013). In plants, senescence is a highly controlled and active process involved in disintegration and remobilization of valuable resources (Maillard et al., 2015). Senescence is followed by a massive degradation of chlorophyll and by inhibition of photosynthetic processes in leaf including photosystem II (PS II) (Janečková et al., 2018) and photosystem I (PS I) activity (Krieger Liszkay et al., 2015). The clearly visible form of senescence in leaf is yellowing caused by chloroplast pigment-protein complexes (Smart, 1995). Therefore, with the onset of senescence in leaf curtails the economic yield of crop plants to a significant extent. Senescence induced by dark is being used experimentally to study the progress of leaf senescence in an easy manner. Some metal ions like Co²⁺ (Geetha ChanGbberellic acid (GA) delay leaf senescence (Zhang et al., 2013; Li et al., 2014).

dra, 1981), Ni (Mishra and Samal, 1971) Al³⁺ (Sudhan and Murthy, 2001) delay senescence in various crop plants. Plant hormones play a major influencing role in each stage of leaf senescence, including the initiation, progression and terminal phase of senescence.

Over the last few years, the study on delay in the senescence of leaf is being carried out widely with high-interest but the studies related to photosynthetic activities are scanty. Hence biochemical characterization of GA +AlCl₃ helped to elucidate few processes that are occurring.

Materials and Methods

The healthy seeds of wheat were obtained from RARS, Tirupati, Andhra Pradesh, India. The seeds were surface sterilized with 0.1% $HgCl_2$ for 2 min and thoroughly washed with tap water and then with distilled water. The seeds were incubated for 6 h and germinated in petri dishes on filter paper for 3 days. The seedlings were randomly placed in plastic trays and watered daily with quarter strength Hoagland nutrient solution and grown in a growth chamber providing a fluorescent light intensity of 30-35 μ moles m⁻² S⁻¹ at 25±1 °C. Fully expanded 8th day leaf segments (4-5 cm long) were cut from the apical region and used for treatment. To study the combined effect of phytohormone 5 μ M Gibberellic acid (GA) and metalion 20 μ M AlCl₃ (Al³⁺) was used. Leaf segments in test solutions were kept in dark at 25 °C for 24-96 h. During the period of treatment, the test solutions were regularly replaced every 24 h with fresh ones.

Estimation of Chlorophyll and Protein Content

The total Chl and protein content was measured using the method of Arnon (1949). The protein content in the leaf segment was determined using Lowry et al. (1951) method.

Electron Transport and Spectral activities

Thylakoid membranes were isolated according to the procedure similar to that of Saha and Good (1970) as described in Swamy et al. (1995).

The Whole chain electron transport activity(WCE) was measured as O_2 consumption by using Methyl Viologen (MV) as an electron acceptor in the thylakoid membranes. The 2 ml reaction mixture contained reaction buffer 50 mM HEPES-NaOH, (pH 7.5), 100 mM Sucrose, 2 mM MgCl₂ and 5 mM KCl, 0.5 mM MV 1.0 mM sodium -azide and thylakoid membranes equivalent to 40 µg of Chl. PS II catalyzed electron transport assay was measured as (H₂O \rightarrow p-BQ) as O₂evolution in the thylakoid membranes. The 2 ml reaction mixture contains reaction buffer 50 mM HEPES-NaOH, (pH 7.5), 100 mM Sucrose, 2 mM MgCl₂, 5 mM KCl, 0.5 mM freshly prepared p-BQ and thylakoid membranes equivalent to 40 µg of Chl. PS I catalyzed electron transport assay was measured as O₂ consumption. The 2 ml reaction mixture contains reaction buffer 50 mM HEPES-NaOH, (pH 7.5), 100 mM Sucrose, 2 mM MgCl₂, 5 mM KCl, 0.5 mM KCl, 0.1 mM 2,6-dichlorophenol indophenols (DCPIP), 0.5 mM MV, 5 mM ascorbate, 1 mM sodium azide, 10 µM DCMU and thylakoid membranes equivalent to 40 µg of Chl.

Polypeptide Analysis

Polypeptide analysis of thylakoid membranes was made according to Laemmli (1970) using SDS-PAGE mini gel apparatus.

Statistical Analysis

All the treatments data are represented as mean \pm SE of five replications. Students T-test was performed to identify the time points at which the mean for GA and corresponding control values are considered significant at p< 0.01.

Results and Discussion Chlorophyll and Protein Content

Total Chl steadily declined to 31% in control leaves segments at 96 h dark incubation. The combined application of GA+ $AlCl_3$ significantly reduced this loss to 48% at 96 h (p< 0.05) (Fig. 1). Total protein content steadily decreased to 34% in control leaves segments at 96 h during dark incubation (Fig. 2). The combined application of GA+ $AlCl_3$ significantly reduced this loss to 79% at 96 h (p< 0.05). Degradation of chlorophyll is a signal in the process of senescence (Krieger –Liszkay et al., 2019). Our results were supported by Doorn et al. (2013) in restoration of protein and chlorophyll content in iris flower senescence by using cytokinins and jasmonates. Reduction of protein and chlorophyll loss by Al^{3+} indicates valency dependent protection by metal ions.







dark-induced senescence. Each value is mean± SE of five replications.

Electron Transport Activities

To relate the persistants of pigments and proteins by GA + AlCl₃ to photochemical activities of thylakoid membranes, WCE, PS II and PS I were assayed. The WCE activity was measured by using MV as an electron acceptor. In the control thylakoid membranes WCE decreased to 42% at 72 h, while the activity was not found at 96 h dark incubation (Table 1). The combined application of GA+ AlCl₃ significantly reduced the WCE loss to 75% at 72h (p< 0.01). p-BQ supported control PS II activity decreased to 38% during dark incubation at 96 h and this loss was reduced significantly to 62% by GA+ AlCl₃ at 96 h (Table 1) (p< 0.01). PS I activity slightly decreased to 78% in control thylakoid membranes at 96h. The combined application of GA+ AlCl₃ reduced this loss to 89% at 96 h (Table 1) (p< 0.01). Our results were supported by Sudan and Murthy (2001) in restoration of PS II, PS I and WCE by combined application of kinetin and aluminium in wheat. In cucumber cotyledons, GA and kinetin influence the functional site of PS I and PS II reaction centers, there-by encouraging the development of the photosynthetic electron transport system (Pedhadiya et al., 1987). Similarly, in broad bean protoplasts, short-term GA-3 treatment increased the net photosynthetic rate and O₂ evolution (Yuan and Xu, 2001).

Photosynthetic activity	Treatment	Incubation time (h)				
		0	24	36	72	96
WCE	Control	115±2	91±2	73±3	48±3	-
		(100)	(79)	(63)	(42)	
	GA+ AlCl ₃	115±2	110±5	99±4	85±7	-
		(100)	(96)	(86)	(75)	
PS II	Control	190±4	170±11	151±4	91±4	73±8
		(100)	(89)	(79)	(48)	(38)
	GA+ AlCl ₃	190±4	188±15	175±12	151±12	118±11
		(100)	(99)	(92)	(79)	(62)
PS I	Control	480±10	447±16	430±11	399±12	399±11
		(100)	(93)	(90)	(83)	(78)
	GA+ AlCl ₃	480±10	470±8	459±15	448±16	425±69
		(100)	(98)	(96)	(93)	(89)

Table 1: Effect of GA+ AlCl₃ on WCE [μM (O₂ consumed) mg⁻¹ Chl h⁻¹], PS II [μM (O₂ evolved) mg⁻¹ Chl h⁻¹] and PS I [μM (O₂ consumed) mg⁻¹ Chl h⁻¹] activities in wheat primary leaf segments under dark incubated senescence.

Each value is mean± SE of five replications.

Values in parenthesis indicate % residual activities.

Spectral Activities

The chlorophyll a absorption and fluorescence derived parameters can be suggested as tool to monitor the leaf senescence. In this connection absorption and fluorescence activities were assayed. Absorption spectra of 0h control thylakoid membranes showed two prominent peaks at 680 nm and 440 nm for the absorption of Chl a and humps at 650 and 480 nm for Chl b and carotenoids respectively. At 72h drastic suppression of peaks took place in control thylakoid membranes (Fig. 3). The suppression of peak heights at 680 nm without being shifted to either side of the spectra and suppression of humps at 480 and 440 nm at 72 h were marginalized in GA+ AlCl₃ treated thylakoid membranes. The above finding suggests an alteration in the primary photochemistry of PS II at 72 h is responsible for the decrease of fluorescence emission ratio in both situations with and without DCMU.

Compared to 0h control, 72h thylakoid membranes showed the loss in fluorescence emission (Fig. 4). GA+ AlCl₃ treated thylakoid membranes reduced this loss at 72 h. This trend was observed in the both situations i.e, with and without DCMU. The ratio of Chl a fluorescence emission in both the situations i.e, with and without DCMU at 0h control thylakoid membrane is 1.67 whereas this value

is decreased to 1.15 in 72 h control thylakoid membranes during dark incubation. GA+ AlCl₃ reduced this loss in fluorescent emission to 1.60 at 72 h (data not shown).



Figure 3: Effect of GA+ AlCl₃ on room temperature absorption spectra of thylakoid membranes in wheat primary leaf segments under dark-induced senescence.





Polypeptide Analysis

In SDS-PAGE polypeptide analysis of control thylakoid membrane, polypeptides degrade at 72 h during dark incubation. However, they were protected from degradation by $GA+AlCl_3$ at 72 h (Fig. 5). Polypeptides in the region of 68 KDa protected by $GA+AlCl_3$ at 72 h, restored the loss in Ps I activity. Polypeptides with molecular weight 43, 33, 23 and 17 KD appeared to be degraded in 72 h dark incubated control leaf thylakoid membrane as the intensity of bands in this region decreased. They were protected by $GA+AlCl_3$ at 72

h of dark incubation (Lane 4). Since GA+ AlCl₃ caused retention of protein in the thylakoid membrane, they can be seen as increased intensity of bands in Lane-4.





Conclusion

In conclusion it was found that the GA+ AlCl₃ reduced the loss of pigments, proteins, electron transport activities, spectral properties. The restoration of whole chain electron transport activity by GA+ AlCl₃ was closely associated with the restoration of PS II activity compared to that of PS I. GA+ AlCl₃ treated leaf thylakoid membranes showed an increase in absorption at 680 nm moderate increase at 480 nm and 440 nm at 72 h during dark incubation. GA+ AlCl₃ protected the degradation of water oxidation complex polypeptides (33, 23, 17 KDa) of PS II and slightly protected the 68 kDa of PS I polypeptides. In combination GA+ AlCl₃ restored the photochemical activities and stabilized the thylakoid membranes during dark incubated senescence.

References

- 1. Arnon DI. "Copper enzymes in isolated chloroplasts". Poly phenol oxidase in Beta vulgaris. Plant Physiol 24 (1949): 1-15.
- 2. Doorn WG., et al. "Delay of Iris flower senescence by cytokinins and jasmonates". Physiol Plant 148.1 (2013): 105-120.
- 3. Geeta Chandra Reddy KS and Mohan Ram HY. "Extension of vase life of cut marigold and Chrysanthemum flowers by the use of cobalt chrolide". Indian J. Exp. Biol 19 (1981): 150-154.
- 4. Janečková H., et al. "The interplay between cytokinins and light during senescence in detached Arabidopsis leaves". Plant Cell Environ 41.8 (2018): 1870-1885.
- 5. Krieger-Liszkay A. "Generation of reactive oxygen species in thylakoids from senescing flag leaves of the barley varieties Lomerit and Carina". Planta 241.6 (2015): 1497-508.

34

- Krieger-Liszkay A, Krupinska K and Shimakawa G. "The impact of photosynthesis on initiation of leaf senescence". Physiol. Plant 166 (2019): 148-164.
- 7. Laemmli UK. "Cleavage of structural proteins during the assembly of the head of bacteriophage T4". Nature 227 (1970): 680-685.
- 8. Li F, Chung T and Vierstra RD. "Autophagy-related II plays a critical role in general autophagy- and senescence-induced mitophagy in Arabidopsis". Plant Cell 26 (2014): 788-807.
- 9. López-Otín C., et al. "The hallmarks of aging". Cell 153 (2013): 1194-1217.
- 10. Lowry OH., et al. "Protein measurement with the poly phenol reagent". J. Biol. Chem 193 (1951): 265-275.
- 11. Maillard A., et al. "Leaf mineral nutrient remobilization during leaf senescence and modulation by nutrient deficiency". Front. Plant Sci 6 (2015): 317.
- 12. Pedhadiya MD, Vaishnav PP and Singh YD. "Development of photosynthetic electron transport reactions under the influence of phytohormones and nitrate nutrition in greening cucumber cotyledons". Photosynth Res 13 (1987): 159-165.
- 13. Saha S and Good NE. "Products of the photo phosphorelation reactions". J. Biol. Chem 245 (1970): 5017-5021.
- 14. Smart CM., et al. "The timing of maize leaf senescence and characterisation of senescence related cDNAs". Physiol. Plant 93 (1995): 673-682.
- 15. Subhan D and SDS Murthy. "Senescence Retarding Effect of Metal Ions: Pigment and Protein Contents and Photochemical Activities of Detached Primary Leaves of Wheat". Photosynthetica 39.1 (2001): 53-58.
- 16. Swamy PM, Murthy SDS and Suguna P. "Retardation of dark induced invitro alterations in PS II organization of cowpea leaf discs by combination of Ca2+ and benzyladenine". Boil. Plant 37 (1995): 457-460.
- 17. Wu X., et al. "Effects of cytokinin on photosynthetic gas exchange, chlorophyll fluorescence parameters and antioxidative system in seedlings of eggplant (Solanum melongena L.) under salinity stress". Acta Physiologiae Plantarum 34 (2012): 2105-2114.
- 18. Yuan L and Xu DQ. "Stimulation effect of gibberellic acid short-term treatment on leaf photosynthesis related to the increase in Rubisco content in broad bean and soybean". Photosynth. Res 68 (2001): 39-47.
- 19. Zhang H, Mishra D and Samal B. "Interaction of benzimidazole and nickel in delaying senescence of detached rice leaves". Z. naturforsch 26c (2013): 1377.
- 20. Zhang K., et al. "Salicylic acid 3-hydroxylase regulates Arabidopsis leaf longevity by mediating salicylic acid catabolism". Proc Natl Acad Sci USA 110 (2013): 14807-14812.